



**You have downloaded a document from  
RE-BUS  
repository of the University of Silesia in Katowice**

**Title:** Current Perspectives on the Auxin-Mediated Genetic Network that Controls the Induction of Somatic Embryogenesis in Plants

**Author:** Anna M. Wójcik, Barbara Wójcikowska, Małgorzata D. Gaj

**Citation style:** Wójcik Anna M., Wójcikowska Barbara, Gaj Małgorzata D. (2020). Current Perspectives on the Auxin-Mediated Genetic Network that Controls the Induction of Somatic Embryogenesis in Plants. "International Journal of Molecular Sciences" (Vol. 21 (2020), Art. No. 1333), doi 10.3390/ijms21041333



Uznanie autorstwa - Licencja ta pozwala na kopiowanie, zmienianie, rozprowadzanie, przedstawianie i wykonywanie utworu jedynie pod warunkiem oznaczenia autorstwa.



UNIwersYTET ŚLĄSKI  
W KATOWICACH



Biblioteka  
Uniwersytetu Śląskiego



Ministerstwo Nauki  
i Szkolnictwa Wyższego



Review

# Current Perspectives on the Auxin-Mediated Genetic Network that Controls the Induction of Somatic Embryogenesis in Plants

Anna M. Wójcik <sup>\*,†</sup> , Barbara Wójcikowska <sup>\*,†</sup> and Małgorzata D. Gaj

Institute of Biology, Biotechnology and Environmental Protection, Faculty of Natural Sciences, University of Silesia in Katowice, Jagiellońska 28, 40-032 Katowice, Poland; malgorzata.gaj@us.edu.pl

\* Correspondence: anna.wojcik@us.edu.pl (A.M.W.); barbara.wojcikowska@us.edu.pl (B.W.)

† These authors contributed equally to this work.

Received: 25 January 2020; Accepted: 14 February 2020; Published: 16 February 2020



**Abstract:** Auxin contributes to almost every aspect of plant development and metabolism as well as the transport and signalling of auxin-shaped plant growth and morphogenesis in response to endo- and exogenous signals including stress conditions. Consistently with the common belief that auxin is a central trigger of developmental changes in plants, the auxin treatment of explants was reported to be an indispensable inducer of somatic embryogenesis (SE) in a large number of plant species. Treating in vitro-cultured tissue with auxins (primarily 2,4-dichlorophenoxyacetic acid, which is a synthetic auxin-like plant growth regulator) results in the extensive reprogramming of the somatic cell transcriptome, which involves the modulation of numerous SE-associated transcription factor genes (TFs). A number of SE-modulated TFs that control auxin metabolism and signalling have been identified, and conversely, the regulators of the auxin-signalling pathway seem to control the SE-involved TFs. In turn, the different expression of the genes encoding the core components of the auxin-signalling pathway, the AUXIN/INDOLE-3-ACETIC ACIDS (Aux/IAAs) and AUXIN RESPONSE FACTORS (ARFs), was demonstrated to accompany SE induction. Thus, the extensive crosstalk between the hormones, in particular, auxin and the TFs, was revealed to play a central role in the SE-regulatory network. Accordingly, LEAFY COTYLEDON (LEC1 and LEC2), BABY BOOM (BBM), AGAMOUS-LIKE15 (AGL15) and WUSCHEL (WUS) were found to constitute the central part of the complex regulatory network that directs the somatic plant cell towards embryogenic development in response to auxin. The revealing picture shows a high degree of complexity of the regulatory relationships between the TFs of the SE-regulatory network, which involve direct and indirect interactions and regulatory feedback loops. This review examines the recent advances in studies on the auxin-controlled genetic network, which is involved in the mechanism of SE induction and focuses on the complex regulatory relationships between the down- and up-stream targets of the SE-regulatory TFs. In particular, the outcomes from investigations on Arabidopsis, which became a model plant in research on genetic control of SE, are presented.

**Keywords:** auxin; Aux/IAA; auxin signalling; ARFs; LAFL group genes; somatic embryogenesis; transcription factors

## 1. Introduction

The process of somatic embryogenesis (SE) demonstrates the unique developmental capacity of plants for switching on the embryogenic programme of development in somatic cells that have already differentiated. As a result of the embryogenic transition, bipolar structures that resemble zygotic embryos, which are called somatic embryos, are formed. Somatic embryos are able to develop into

complete plants and since 1960s, numerous protocols have been established that enable the efficient regeneration of dozens of plant species via somatic embryos that are induced under in vitro culture conditions (reviewed in [1]). In addition to its wide implementation in the mass micropropagation of plants for the market, the SE-based protocols are also commonly used to produce transgenic plants (reviewed in [2]). In addition to its practical significance in plant biotechnology, the SE process provides a valuable model system for studies on plant embryogenesis and the developmental plasticity of the plant somatic cells [3,4]. Identifying the exo- and endogenous factors that promote embryogenic transition in in vitro-cultured somatic cells leads to a better understanding of plant cell totipotency. In contrast to advanced knowledge on the tissue culture conditions that promote SE induction [5], the endogenous factors that determine the tissue capacity for an embryogenic response are much less recognised. However, distinct progress in deciphering the molecular mechanism that governs the embryogenic transition of plant somatic cells has been made in the last decade and the role of auxin in SE induction has been intensively studied at the molecular level.

## 2. Auxin, a Main Inducer of SE

Since the classical demonstration of SE induction in carrot callus [6], the impact of auxin on the embryogenic pathways that are induced in in vitro-cultured plant tissues has become evident and hundreds of protocols have been established to induce SE in a plethora of plant species [7]. Although there are some differences between the in vitro culture conditions that are applied to induce SE in different plants, treating explants with auxin and auxin-like substances seems to be commonly required for SE induction. In support of this statement, auxin treatment was applied in 74 of the 80 protocols reviewed in Table S1 and auxin was frequently combined with cytokinins (65%). The most effective and commonly used inducer of SE (78% of the protocols) is 2,4-dichlorophenoxyacetic acid (2,4-D), which is a synthetic plant growth regulator of auxin activity.

## 3. Auxin- and Stressor-Like Activity of 2,4-D in SE Induction

2,4-D was initially used in cereal agriculture as a systemic herbicide that selectively kills broadleaf weeds [8]. Treatment with 2,4-D causes a number of morphological changes in plants in a concentration-dependent manner. 2,4-D efficiently stimulates cell division, plant growth and in vitro-induced morphogenetic responses at low concentrations of 1–10 mg/L, while spraying the plants in the field with 2,4-D of a concentration of about one-thousand-fold higher (2 g/L, according to the weed control manuals) caused lethal symptoms that were similar to an auxin overdose [9]. Of note, in spite of the distinctly higher tolerance of the monocot vs. the dicot plants to the toxic effects of 2,4-D [10], similar concentrations of this substance are applied to induce SE in different plants, and relevantly, 1–2 mg/L of 2,4-D is recommended for SE induction in maize and wheat [11–14], while 1.1 mg/L of 2,4-D is effective for establishing an embryogenic culture in Arabidopsis [15].

2,4-D shares structural similarity with indole-3-acetic acid (IAA), which involves the presence of a carboxyl group and an aryl ring structure [10]. However, in contrast to natural auxin, IAA, which is rapidly inactivated via conjugation and degradation in plant cells [16], the effects of 2,4-D on plants are long-lasting because of its high level of stability in the plant cells [17]. Moreover, the polar cell-to-cell transport of 2,4-D and IAA differs in terms of the auxin carriers that are engaged. Accordingly, in contrast to IAA, which is transported polarly by both types of auxin carriers, i.e., the efflux PIN-FORMED (PIN) and P-GLYCOPROTEIN (PGP) and the influx AUXIN1/LIKE-AUX1 (AUX/LAX) proteins, the transport of 2,4-D by the PIN efflux carriers is limited, which results in an accumulation of 2,4-D in plant cells [18]. Moreover, 2,4-D has a weak binding affinity to the IAA receptor, AUXIN-BINDING PROTEIN1 (ABP1) [19]. However, similar to IAA, 2,4-D is able to promote the binding of AUXIN/INDOLE-3-ACETIC ACID (Aux/IAA) proteins to the TRANSPORT INHIBITOR RESPONSE1 (TIR1) F-box protein of the auxin-signalling pathway after the auxin responsive genes are activated by the AuxRE (AUXIN RESPONSIVE ELEMENT) motif in the promoters [20,21]. It is worth noting that the 2,4-D and IAA-induced transcriptomes tend to overlap only to some extent [20],

which reflects the differences between IAA and synthetic auxin in their interactions with the signalling machinery [22].

In addition to its auxin-like activity, 2,4-D is believed to affect the plant developmental processes by inducing the stress responses. Accordingly, treatment with 2,4-D causes an accumulation of ROS (reactive oxygen species) and the stress hormones ethylene and abscisic acid (ABA) in plant tissue [23–25]. Relevant to the stress-related mechanism of 2,4-D activity, the central role of stress factors, in particular, oxidative stress, in promoting the developmental switches in plant cells including the induction of SE has been widely demonstrated (reviewed in [26,27]). In support of the stressor-like activity of 2,4-D, numerous stress-responsive genes were found to be differently expressed in 2,4-D-induced SE-transcriptomes [28–33].

However, a long-lasting debate on the auxin- vs. stress-like activity of 2,4-D seems to be groundless given that the complex relationships between auxin and stress responses in controlling plant development have been documented, and relevantly, different stress factors impact auxin signalling and biosynthesis [34] (reviewed in [35]). Moreover, 2,4-D treatment also activated the core regulators of the auxin-signalling AUXIN RESPONSE FACTORS (ARFs) and the TRYPTOPHAN AMINOTRANSFERASE OF ARABIDOPSIS1/YUCCA, TAA1/YUC-dependent auxin biosynthesis pathway in SE-induced explants of Arabidopsis [36,37]. Thus, both the auxin- and stressor-like intrinsic function of 2,4-D are closely related in promoting embryogenic development in plant cells.

#### 4. Biosynthesis and Accumulation of Auxin During SE

The observation that the embryogenic explants of various plants contain a higher level of IAA than the non-embryogenic explants implies that there is a positive relationship between the auxin content and the capacity of tissue to induce SE [38–45]. However, in conifers, the SE-recalcitrant cultures had a higher level of IAA [46,47], which suggests that the auxin content that enables SE induction seems to be genotype and tissue specific. It seems that not the initial level of IAA in explant tissue but rather a dynamic increase of the endogenous auxin content in response to the SE-induction factors might provide an indispensable signal for the embryogenic transition. Relevantly, a significant increase in the IAA content was reported to accompany early SE induction in different plants including carrot [48], sunflower [49], robusta coffee [50–52], alfalfa [53], pineapple guava [54], Arabidopsis [36], cotton [55] and Norway spruce [56]. Besides in vitro culture conditions, transgenic plants of Arabidopsis that produced somatic embryos in planta as a result of the overexpression of the *LEAFY COTYLEDON2* (*LEC2*) gene were also found to accumulate IAA [57]. Auxin accumulation in plant tissue in response to the SE-induction signal resembles the early stages of ZE. Accordingly, an increase in the endogenous auxin level was observed in the early-stage zygotic embryos of carrot [58] and other plants [54]. In the ZE of Arabidopsis, IAA is maternally biosynthesised de novo in the fertilised ovules and provides a source of auxin for the correct embryo development [59,60]. The auxin accumulation that accompanies the embryogenic development that is induced in both generative and somatic plant cells provides further support for a similarity in the molecular and hormonal signals that control embryogenic development in vivo and in vitro [3,61].

Two alternative metabolic pathways, the tryptophan-dependent and -independent control de novo IAA biosynthesis in plant development including zygotic embryo development [59,62,63]. In an SE culture of Arabidopsis, a TAA1/YUC pathway, which is controlled by the tryptophan aminotransferases TAA1 and TRYPTOPHAN AMINOTRANSFERASE RELATED1-4 (TAR) and YUCCA flavin-dependent monooxygenases, has been identified [36]. Six of eleven *YUC* genes, *YUC1*, *YUC2*, *YUC4*, *YUC6*, *YUC10* and *YUC11*, were found to be expressed in different embryogenic cultures of Arabidopsis [36,64,65]. It is worth noting that the SE-associated *YUCs* were specifically expressed in the shoot part of a plant and that most of them were active in ZE [66,67]. The expression of the SE-involved *YUCs* was localised in the SE-involved explant part, i.e., in the cotyledons of the immature zygotic embryo, and that the relevant single and multiple *yuc* mutants, including *yuc2*, *yuc4* and *yuc1*, *yuc4*, *yuc10* and *yuc11* had a severely reduced capacity for SE [36,65,68]. At the transcriptional level, auxin biosynthesis is regulated

by the developmental and environmental signals that control the binding of specific TFs to the auxin biosynthesis-related genes including the *YUCs* [69]. Thus, the set of SE-involved *YUC* genes might differ among the plants, explants and treatment that is applied. Accordingly, in *Arabidopsis*, the 2,4-D treatment of immature zygotic embryos (IZE) explants activated *YUC1*, *YUC4* and *YUC10* [36], while only two of these genes were stimulated in the developing somatic embryos of the trichostatin A-induced IZE explants [68]. Ethylene was indicated to control the *YUC* expression and the negative impact of ethylene on SE induction in *Arabidopsis* was attributed to the inhibition of the *YUCs*, and subsequently, a disturbed auxin accumulation in the culture [65].

In line with the results on *Arabidopsis*, a RNA-seq analysis of the SE-transcriptome in cotton revealed the up-regulation of numerous genes that are involved in the tryptophan-dependent auxin biosynthesis, including the *YUCs*, *TRP1* (*TRYPTOPHAN BIOSYNTHESIS1*), *ASB1* (*ANTHRANILATE SYNTHASE*), *TSB2* (*TRYPTOPHAN SYNTHASE B-SUBUNIT2*), *NIT4* (*NITRILASE4*) and *CM1* (*CHORISMATEMUTASE*) [70,71]. An increase of the *YUC1* and *TAA1* transcripts that correlated with an increased level of endogenous IAA was also observed in the embryogenic explants of coffee [50]. Thus, the tryptophan-dependent TAA1-*YUC* pathway of IAA biosynthesis seems to commonly contribute to the auxin accumulation that is associated with the embryogenic transition in the somatic cells of plants. However, insights into the available RNA-seq data of an *O. sativa* embryogenic culture [72] indicated only a limited number of auxin biosynthesis-related transcripts and suggested a much less intensive activity of the TAA1-*YUC* IAA biosynthesis pathway in this plant than the one in *Arabidopsis* [73]. However, understanding the auxin biosynthesis pathways in *O. sativa* is still not well developed and further analyses are needed to reach a conclusion about the SE-associated auxin production in the model plants *Arabidopsis* vs. *O. sativa*.

## 5. The Core Regulatory Components of the Auxin-Signalling Pathway That Are Involved in SE

A key insight from the analyses of the SE-related transcriptomes in numerous plants, including wheat [74], palm [32], cotton [70,71,75], *Arabidopsis* [64,76], camphor tree [77], mangosteen [78], cork oak [79], Indian bean tree [80], coffee [81], maize [11] and lily [82], is that besides auxin biosynthesis other aspects of auxin action including auxin perception, polar transport and response/signalling are also involved in the mechanism of SE induction.

The core components of the auxin-signalling pathway that translate auxin sensing into transcriptional responses include the SKP/CULLIN/F-BOX-ubiquitin ( $SCF^{TIR1}$ ) complex, the Aux/IAA transcriptional co-regulators and the sequence-specific binding proteins, which are called ARFs. The  $SCF^{TIR1}$  complex comprises TIR1, the AUXIN-SIGNALLING F-BOX1 (AFB1), AFB2 and AFB3 proteins, which are members of the clade of the auxin receptors family (TAARs). TAARs are a component of the SCF-ligase complexes, which direct the members of the AUX/IAA transcriptional repressor protein family to a proteasome-dependent degradation. The degradation of AUX/IAA releases the TOPLESS (TPL) transcriptional co-repressor and allows the ARF proteins to bind at the promoters of the primary auxin responsive genes in order to regulate their transcription [83]. Importantly for the role of auxin in establishing the SE-transcriptome in the explant cells, the specific expression pattern of the auxin-responsive genes is likely due to the specific cellular concentrations of several of the Aux/IAA proteins that seem to result from the auxin content in a cell. However, a quantitative relationship between cellular auxin concentration and gene activity has not yet been indicated, and whether there are separable gene sets that only respond to low or high auxin concentrations remains to be determined [69].

Auxin regulates the specific interactions between the Aux/IAAs and ARFs and almost all of the Aux/IAA proteins can interact with a subset of the ARFs with a long, Q-rich middle region (activating ARFs), whereas interactions with other ARFs are both limited and more specific [84]. In *Arabidopsis*, 29 and 23 members of the *AUX/IAA* and *ARF* gene families have been identified, respectively, and specific AUX/IAA-ARF interactions are assumed to control the auxin-dependent developmental processes including ZE [84,85]. Therefore, identifying the spatiotemporal and



cell-specific sets of the AUX/IAA elements and the interacting ARFs that contribute to the embryogenic transition of somatic cells is required for deciphering auxin-mediated SE induction.

The transcripts encoding AUX/IAAs and ARFs are highly represented in the SE-transcriptomes of Arabidopsis [76,86] and other plants, including cyclamen [87], wheat [74], palm [32,88], cotton [71], camphor tree [77], mangosteen [78], cork oak [79], Indian bean tree [80], coffee [81], maize [11] and lily [82]. Insight into the SE-transcriptomes revealed that the expression profiles of the AUX/IAA and ARF genes are highly diverse and that the same gene might show both a significant up- and down-regulation depending on the embryogenic system (Table S2). The high degree of the diversity of the expression patterns of the ARF genes reflect the complexity of the transcriptional regulation of these genes, which are fine-tuned by numerous factors including the hormonal status of a tissue [84]. The direct regulation of the SE-expressed ARFs by auxin might be considered and an auxin-responsive AuxRE module was found in the majority (84%) of the SE-modulated ARFs in Arabidopsis, including ARF5, ARF6, ARF8, ARF10 and ARF16 [37].

In contrast to the numerous SE-transcriptomic data, functional analyses of the specific auxin-signalling genes that are engaged in SE are still rare and limited almost exclusively to Arabidopsis. A major drawback for such an analysis seems to be the high functional redundancy within members of the TAAR, AUX/IAA and ARF families and thus, the frequently unobvious and pleiotropic phenotype of the relevant mutant plants [84,85]. Analysis of plants with impaired TAAR receptors of auxin, the *tir1-1* and *afb2-3* mutants, indicated their hyposensitivity to 2,4-D treatment and a reduced embryogenic response in vitro and the *TIR1* and *AFB2* genes were also postulated to control SE induction via an miR393-mediated mechanism [89]. In addition to Arabidopsis, the down-regulation of *TIR1* and *AFB2* was also observed during SE in cotton but the regulatory pathway that controls their expression has not yet been identified [70,75,90]. Considering that the TAAR proteins have a different binding affinity to auxin herbicides [91], the high binding affinity of the SE-involved *TIR1* and *AFB2* to the 2,4-D that is used for SE induction in Arabidopsis might be expected [89].

The mutations in the *IAA16*, *IAA29*, *IAA30* and *IAA31* genes that resulted in an impaired embryogenic response of the Arabidopsis explants suggests the engagement of these genes in SE induction [76,86]. The expression of the SE-involved AUX/IAA genes seems to be regulated by the TFs that have a master function in the embryogenic transition, and relevantly, AGAMOUS-LIKE15 (*AGL15*) controls the expression of *IAA16* and *IAA30*, while *LEAFY COTYLEDON2* (*LEC2*), which interacts with *AGL15* in a feedback regulatory loop (see next paragraph), directly targets *IAA30* [86,92].

In accordance with SE-transcriptome data that suggest that the majority of the ARF genes seem to be engaged in controlling SE (Table S2), numerous *arf* mutants *arf1-3*, *arf3*, *arf5-8*, *arf6*, *arf7*, *arf8* were found to be impaired in their embryogenic response [37]. It is noteworthy that the mutants in the ARF genes of the contrasting SE-expression profiles were found to display a reduced embryogenic response, and accordingly, the mutations in *ARF5/ARF6* and *ARF3* with an up- and down-regulated expression in SE, respectively, were indicated to negatively affect SE induction [37]. The lack of a direct relationship between the ARF gene expression profile and the phenotype of the relevant *arf* mutant implies the complexity of the factors that fine tune the ARF-mediated responses during SE induction. These factors include the distinct spatio-temporal and hormone-dependent regulation of ARF gene expression and the complex regulatory interactions within the ARF family members and between the ARFs and other components of the auxin-signalling pathway [37,84].

Several pieces of evidence suggest that similar to ZE (reviewed in [93]), *ARF5*, which encodes the MONOPTEROS (MP) protein, might have a fundamental role in regulating different auxin-controlled aspects of SE. Accordingly, *ARF5* was observed to be the most highly up-regulated member of the ARF family during SE induction and the *arf5* mutant had a significantly reduced capacity for an embryogenic response [37]. One of the functions of *ARF5* in SE seems to be connected with regulating *PHABULOSA* (*PHB*), which is a positive regulator of *LEC2* in ZE [94] and in support of this assumption, in contrast to the up-regulated expression of *PHB* and *LEC2* in a WT embryogenic culture, an *arf5* mutant culture displayed a significantly reduced *LEC2* transcript level [37,95]. A regulatory *ARF5*-*PHB* connection was

observed in vivo [96]. Thus, the ARF5 protein was postulated as controlling the TAA1-YUC-mediated pathway of auxin biosynthesis that operates in SE induction by regulating *PHB* and *LEC2* [36]. In line with this hypothesis, ARF5 targets the components of this pathway (*TAA1* and *YUC1*, *YUC8*) in order to transcriptionally initiate the embryonic ground tissue in an early zygotic embryo [97]. ARF5 might also contribute to the in vitro-induced embryogenic development by controlling the other *TF* genes that have a documented activity in SE, including *HOMEODOMAIN GLABRA 8* (*ATHB8*) and *TARGET OF MONOPTEROS 3* (*TMO3*), *TMO5*, *TMO6* and *TMO7* [64,76]. In support of this assumption, the regulatory relationships between these TFs and ARF5 were identified in planta [98,99]. Interestingly, the *TMO7*, which is a direct ARF5 target encoding a small TF, moves intercellularly to control the embryonic root specification [99]. Hence, it is tempting to question whether ARF5 also controls the mobility of *TMO7* during SE induction and how the ARF5-*TMO7* regulation could contribute to establishing the embryonic identity in the somatic cells of an explant.

Many members of the Aux/IAA family have been shown to interact with ARF5 in the development of Arabidopsis in planta [100] and among them is IAA30 of indicated involvement in SE [76,92]. The possible function of ARF5 may also involve regulating the *PIN* genes, which control the polar auxin transport during in vivo development [101,102] and in support of this statement, a *pin1-7* mutant had a reduced embryogenic induction in vitro [103] and the 35S::ARF5 line, which phenocopies the *pin1* mutant [104], was found to be incapable of inducing SE [37]. In order to stimulate the expression of *PIN1* and to control early zygotic embryo patterning, ARF5 cooperates with the WUSCHEL-RELATED HOMEODOMAIN (WOX) TF gene family members, including *WOX1*, *WOX2*, *WOX5*, *WOX8* and *WOX9* [105–107]. Although the expression of the *WOX2*, *WOX8* and *WOX9* genes was reported in an embryogenic culture of several plants, including grapevine, spruce, Arabidopsis, cotton, barrelclover, larch and longan [64,75,108–114], the auxin-signalling regulators that control the *WOX* genes in SE induction have not been studied as yet. In particular, *WOX2* seems to be an interesting ARF5 target candidate in regulating SE because of its role in balancing the cytokinin versus the auxin hormone pathways during the establishment of the shoot identity in the zygotic embryo [115]. Relevantly, *ARF10*, which has a positive regulatory impact on de novo shoot regeneration via the activation of the shoot meristem-specific genes, was highly expressed in the SE of Arabidopsis [37,116]. Therefore, *ARF10* together with *ARF16* and *miR160* have been postulated to contribute to the *LEC2*-controlled auxin biosynthesis in SE-induced tissue [95]. Besides *LEC2*, other SE-related candidate targets of *ARF10/ARF16* involve the *PLETHORA* (*PLT*) genes, which have an essential function in integrating the hormonal inputs during many developmental processes in plants including the auxin responses in the early embryogenic induction (reviewed in [117]). Because *ARF10* and *ARF16* act upstream of the *PLTs* to regulate the stem cell differentiation in Arabidopsis roots [118], the significant accumulation of *ARF10* and *ARF16* transcripts was accompanied by an increased expression of *PLT1* and *PLT2* in an embryogenic culture of Arabidopsis [37,64,76].

Data on *ARF* expression profiling in the SE of Arabidopsis and other plants also suggests that *ARF6*, *ARF7*, *ARF8*, *ARF9* and *ARF19* (Table S2) play a role in controlling SE induction; however, their specific functions and the SE-involved targets have not yet been identified.

## 6. Complex Interactions Between the *TF* Genes that Control Auxin-Induced SE

The transcripts of the *TF* genes that are related to hormone responses, in particular those that control auxin metabolism and signalling are overrepresented in the SE-transcriptomes of many plants (reviewed in [119]). In Arabidopsis, over 40% of the SE-modulated *TF* genes have been annotated to hormone-related pathways, primarily to the auxin metabolism and signalling pathways [76]. The best recognised TF-regulatory network that controls SE induction includes the *LAFL* group of genes (for *LEC1/L1L*, *ABSCISIC ACID INSENSITIVE 3* (*ABI3*), *FUSCA3* (*FUS3*) and *LEC2*), which have a major and redundant regulatory function in controlling the identity and maturation of a zygotic embryo [120,121]. The overexpression of two of the *LAFL* genes, *LEC1* and *LEC2*, in Arabidopsis seedlings resulted in somatic embryo formation, while the *lec1* and *lec2* mutations significantly inhibited

the embryogenic response of the in vitro cultured explants [122–124]. *LEC1* and *LEC2* gene expression is co-localised with the SE-involved parts of the 2,4-D-treated explants [125–127]. Importantly, *LEC1* and *LEC2* seem to have a common SE-promoting function in plants and the overexpression of these TFs has been suggested as an efficient method for improving the embryogenic response in several crop species, including cassava, rapeseed, tobacco and cacao [128–131]. Taken together, these observations led to a conclusion on universal and essential function of *LEC1/LEC2* in controlling both the somatic and zygotic embryonic development in higher plants (reviewed in [132]).

The diversity of the *LEC1* and *LEC2* targets implies that these proteins contribute to the regulation of various auxin-related processes, which involve the control of auxin biosynthesis (see the paragraph ‘Biosynthesis and Accumulation of Auxin During SE’) and relevantly, members of the Aux/IAA family were postulated to be under the control of *LEC1* (*IAA5*, *IAA16* and *IAA19*) and *LEC2* (*IAA1*, *IAA17*, *IAA30* and *IAA31*) during embryonic development [92,133,134]. The impact of *LEC2* on auxin polar transport also cannot be ruled out because the upregulation of the auxin efflux facilitators, *PIN1* and *PIN2*, was observed in the transgenic tobacco plants that over-expressed *LEC2* [129]. Moreover, the auxin-responsive AuxRE motif, which is present in the *LEC1* and *LEC2* promoters, is evidence of the involvement of the ARF proteins in controlling the auxin-stimulated expression of these genes [37]. The candidate ARFs that control *LEC1/LEC2* in the SE-induced explants might be searched for among the ARFs that have an up-regulated expression in embryogenic cultures (Table S2).

In addition to their auxin-related function, the *LEC* genes (*LEC1* and *LEC2* together with *FUS3*) seem to control the ability of explants to respond to SE via the control of the gibberellins (GA)/ abscisic acid (ABA) balance in tissue. Accordingly, *AGL15*, which is an activator of the *GA2ox6* gene that encodes an enzyme that inactivates bioactive gibberellin, was identified among the direct targets of *LEC2* [92,132,135]. *AGL15* encodes a MADS box TF that increases the embryogenic competency of tissue when it is ectopically expressed [136]. The expression of *AGL15* is autoregulated [137] and regulated by feedback interactions with other TFs, which involve the direct regulation of *AGL15* by *LEC2* and *FUS3* and the control of *LEC2* and *FUS3* by *AGL15* [86,92,138,139].

Insights into the *AGL15*-overexpression transcriptomes of Arabidopsis and soybean showed that *AGL15* might enhance the embryogenic competency of the explants by promoting tissue dedifferentiation [140] and also revealed the complex interactions of *AGL15* with the hormone pathways including auxin signalling [141]. Accordingly, *AGL15* was demonstrated to negatively regulate the auxin-signalling components, including the direct repression of *ARF6* and *TIR1* ubiquitin ligase, the indirect repression of *ARF8* and the direct up-regulation of *IAA30* [86,141].

The SE-regulatory network of TFs also involves members of the AINTEGUMENTA-LIKE (AIL) family of the APETALA2/ETHYLENE RESPONSE FACTOR (AP2/ERF) domain TFs [117]. The small AIL family comprises eight members, including the AINTEGUMENTA (ANT), AIL, BABY BOOM (BBM/PLT4) and five other PLT proteins that have overlapping functions in the dividing cells of meristematic and embryonic tissue [142,143]. Of all of the AIL TFs, a central position in regulating SE has been attributed to BBM/PLT4; however, other PLTs (PLT1, PLT2, PLT3, EMK/PLT5 and PLT7) also promote embryogenic induction in response to its overexpression [121,144,145]. Recently, PLT1/PLT2 was reported to control the *LEC2* function in auxin-induced SE and the miR396–GRF module has been postulated to control the PLT2-*LEC2* interaction [146].

To induce SE, *LEC1* and *LEC2* are positively regulated by BBM, which is another major TF protein that controls the identity and totipotency of the plant embryo, and the SE-promoting activity of BBM requires the LAFL/AGL15 proteins [147]. Knowledge about the other functionally characterised targets of BBM is limited. In search of direct BBM targets, a set of mostly uncharacterised genes that have assumed functions in transcription, cellular signalling and cell wall biosynthesis was identified including the *ACTIN DEPOLYMERISING FACTOR9* (*ADF9*) gene, which encodes an ADF/cofilin protein [148]. Auxin-related genes were revealed within the BBM targets that are involved in SE in Arabidopsis including the genes that are associated with the biosynthesis (*TAA1*, *YUC3* and *YUC8*), transport (*PIN1* and *PIN4*) and signalling of auxin (*ARF2*, *ARF6*, *ARF10*, *IAA2*, *IAA7* and *IAA28*) [121].



In addition, a possible role of PLTs in the auxin-related mechanism of SE induction implies the activation of the *YUC* genes (*YUC1* and *YUC4*) by PLT3, EMK/PLT5 and PLT7 in order to stimulate auxin biosynthesis in phyllotaxis [149].

The activators of the LEC-mediated SE pathway also involve the class III HOMEODOMAIN LEUCINE ZIPPER (HD-ZIP III) TFs, PHB and PHAVOLUTA (PHV), which activate *LEC2* in the vegetative development of Arabidopsis [94]. Similarly, PHB/PHV and *LEC2* might interact in SE through a positive feedback loop, which, under the repression of miR166, was postulated to control the YUC-pathway of auxin biosynthesis in explants that were undergoing embryogenic induction [95].

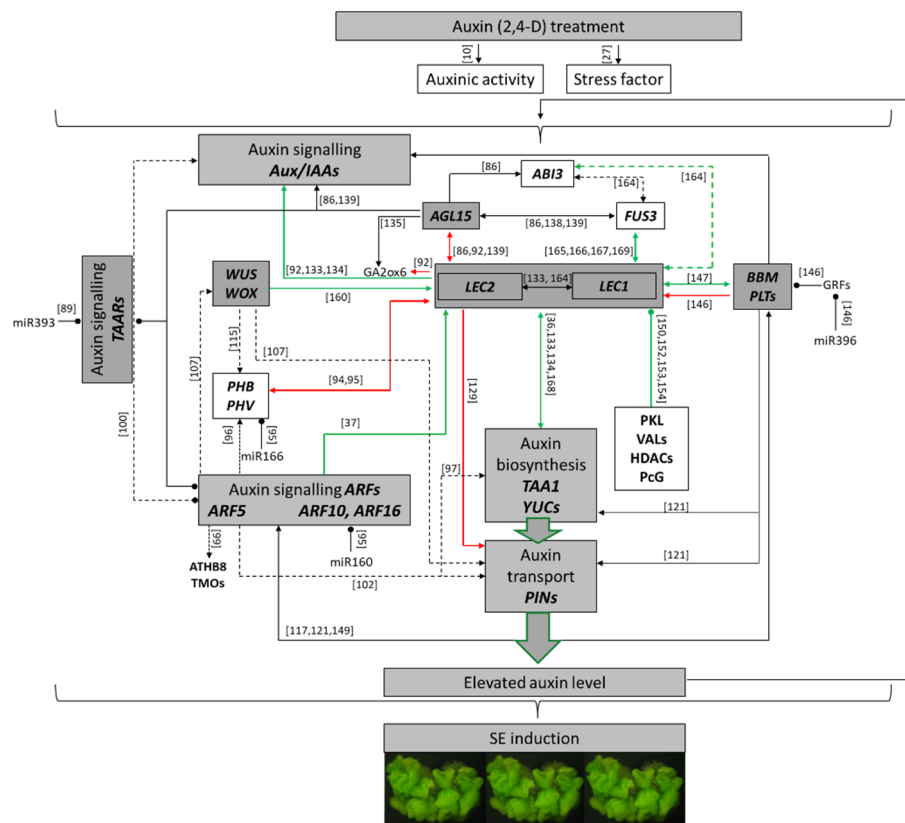
Several TFs have been recognised to negatively control the *LAFL* genes. The PICKLE (PKL) protein of the CHD3-type ATP-dependent SWI/SNF chromatin-remodelling factors impacts the global H3K27me3 levels including *LEC1* and *LEC2* [150,151]. In addition, the VIVIPAROUS1/ABI3-LIKE (VAL)/HIGH-LEVEL (VAL1) and VAL2 proteins that like ABI3, FUS3 and *LEC2* have a B3 domain were postulated to function as global regulators of the *LEC1/B3* gene system, which controls embryo identity in seedlings [152]. The interactions of the VALs with the chromatin factors and the gene repression mechanism that involves the VAL-mediated recruitment of HDAC histone deacetylases and Polycomb group proteins to the target loci were recognised [153,154].

An SE-promoting capacity was also indicated for *WUSCHEL* (*WUS*), which is a member of the *WOX* superfamily of the genes encoding the plant-specific homeobox TFs that are involved in promoting cell division and/or preventing the premature cell differentiation of the stem cells in SAM (reviewed in [155,156]). The positive impact of *WUS* on the embryogenic potential of tissue was indicated given that the gain-of-function mutation in *WUS* led to the spontaneous formation of somatic embryos on Arabidopsis root explants [157] and an overexpression of *WUS* promoted or enhanced the embryogenic response in other plants [158–160]. Auxin treatment was found to be essential for correctly regulating the *WUS* expression and the *WUS* transcripts that were detected in a group of embryogenic callus cells were postulated to indicate an auxin accumulation in the SAM and the pro-embryo [103]. Although the ectopic expression of *AtWUS* in transgenic cotton callus was associated with the upregulation of the auxin biosynthesis-regulators, the *LEC1* and *LEC2* genes [160], the content of IAA was not modulated [161]. Hence, the contribution of *WUS* to the TF-regulatory network of SE seems to be unrelated to auxin biosynthesis and in support of this assumption, within the numerous auxin-related targets of *WUS*, the genes that are involved in auxin biosynthesis have not been identified [162]. In the mechanism of the *WUS*-mediated control of stem cells in SAM, *WUS* acts as an auxin rheostat that controls the target loci including numerous genes of the auxin-signalling pathway by regulating histone acetylation [162].

Insights into ZE regulation suggest that in addition to *WUS*, other members of the *WOX* genes family might also interact with the LEC-controlled pathway of SE induction. Accordingly, during stem cell initiation in early zygotic embryo, the *WOX* genes (*WOX2* and its paralogs *WOX1*, *WOX3* and *WOX5*) were shown to positively control the PHB/PHV TFs that activate the *LEC* genes in both zygotic and somatic embryogenesis [95,115]. Similar to ZE, the regulatory interaction between the *WOX*s and PHB/PHV might be expected during somatic embryo formation because the expression of both *WOX5* and *PHB/PHV* was indicated in the embryogenic and callus cells of Arabidopsis [163]. Moreover, the *WOX2*, *WOX8* and *WOX9* transcripts accompany different stages of somatic embryo development in different plants [64,75,108–114]. However, the regulatory interactions between the *WOX* genes and the LEC-controlled SE pathway require experimental validation.

## 7. Conclusions and Future Prospects

In the past ten years, omics data analysis together with the candidate gene approach have significantly contributed to identifying the regulatory interactions between the TFs that have a key function in SE and the phytohormones, mainly auxin. A revealing picture of the SE-regulatory gene network reveals a high level of the complexity of the extensive crosstalk between the SE-TFs and auxin during the auxin- induced embryogenic development in somatic plant tissue (Figure 1).



**Figure 1.** An overview of the regulatory interactions that control SE induction. Treating explants with auxin (mostly 2,4-D) activates a number of the TF genes that have a regulatory function in embryonic development, including the *LEC1* and *LEC2* of the LAFL group, *BBM* (*PLTs*), *AGL15* and *WUS/WOX* genes. The down-stream targets include the genes of the TAA1/YUC of the auxin biosynthesis pathway, the PINs that control auxin transport and the core regulators of auxin signalling (AUX/IAAs and ARFs). Other regulatory elements of the SE-network also involve miRNAs and the epigenetic regulators (PKL, VALs, HDACs). The arrows and circle-shaped ends indicate the activation or repression of gene expression, respectively. The solid and dashed lines indicate the experimentally validated and suggested regulatory interactions, respectively. The red and green lines indicate the regulatory interaction between LEC2 and LECs, respectively [164–169].

Significant progress in establishing the genetic network that controls SE has been observed and both the down-stream targets and up-stream regulators of the SE-involved TFs have been identified and their functions in relation to auxin have been revealed. However, knowledge about the role of auxin in the regulatory network that controls SE induction is still fragmentary since in addition to gene transcription, auxin regulates a plethora of other cellular processes, including the epigenetic state of chromatin, protein stability/protein-protein interactions and metabolic pathways. Moreover, the interactions of auxin with other hormone pathways, in particular cytokinins, need to be explored in cells that are undergoing SE induction. Since only a small subset of explant cells is capable of embryogenic transition, the methods that enable a single-cell resolution analysis would provide new insights into the auxin-mediated regulatory network that governs the unique developmental plasticity that is demonstrated by somatic plant cells.

The ongoing debate on the SE regulatory network also concerns the degree of the molecular similarity of the embryogenic development that is induced in vitro in somatic tissue to the ZE process. In accordance with the common belief of a close similarity of the regulatory pathways that control both types of plant embryogenesis, several master regulators of ZE, including LEC, PLT and WOX TFs, were found to play a decisive role in the embryogenic reprogramming of plant somatic cells. Moreover, the auxin-related processes including auxin biosynthesis and signalling was placed in centre of SE and

ZE regulation. However, recent RNAseq data of an embryogenic culture of *Arabidopsis* and *Pinus pinaster* showed that the SE-transcriptome seems to be distinctly different from the transcriptome of a zygotic embryo [170,171]. This unexpected finding suggests that the molecular events that trigger SE induction might differ from the ones that operate during ZE and points to the epigenetic processes that orchestrate the transcriptomes of in vitro cultured somatic cells in response to plethora of endo- and exogenous factors, thus making them different from those found in ZE. Therefore, identifying the epigenetic processes that contribute to the embryogenic transition in somatic plant cells in response to auxin would lead to further progress in revealing the SE regulatory network.

**Supplementary Materials:** The following are available online at <http://www.mdpi.com/1422-0067/21/4/1333/s1>.

**Author Contributions:** All of the authors listed have made a substantial, direct and intellectual contribution to the work and approved it for publication. All authors have read and agreed to the published version of the manuscript.

**Funding:** This work was supported by a research grant from the National Science Centre in Poland (OPUS13 2017/25/B/NZ1/01615).

**Conflicts of Interest:** The authors declare no conflict of interest.

## Abbreviations

2,4-D	2,4-Dichlorophenoxyacetic acid
AFB	AUXIN F-BOX PROTEIN
AGL15	AGAMOUS-LIKE15
ARF	AUXIN RESPONSE FACTOR
Aux/IAA	AUXIN/INDOLE-3-ACETIC ACID
AuxRE	AUXIN RESPONSIVE ELEMENT
BBM	BABYBOOM
HDAC	HISTONE DEACETYLASE
IAA	indole-3-acetic acid
IZE	immature zygotic embryos
LAFL	LEC1, ABI3, FUS3, LEC2
LEC	LEAFY COTYLEDON
miRNA	microRNA
PIN	PIN-FORMED
PHB	PHABULOSA
PHV	PHAVOLUTA
PLT	PLETHORA
SAM	shoot apical meristem
SE	somatic embryogenesis
TAA1	TRYPTOPHAN AMINOTRANSFERASE OF ARABIDOPSIS1
TAAR	TIR1/AFBs clade of Auxin Receptors
TAR	TRYPTOPHAN AMINOTRANSFERASE RELATED
TF	transcription factor
TIR1	TRANSPORT INHIBITOR1
YUC	YUCCA
WOX	WUSCHEL-RELATED HOMEBOX
WUS	WUSCHEL
ZE	zygotic embryogenesis

## References

1. Monja-Mio, K.M.; Herrera-Alamillo, M.Á.; Robert, M.L. Somatic embryogenesis in temporary immersion bioreactors. In *Somatic Embryogenesis: Fundamental Aspects and Applications*, 1st ed.; Loyola-Vargas, V., Ochoa-Alejo, N., Eds.; Springer International Publishing: Cham, Switzerland, 2016; pp. 435–454.

2. Ochoa-Alejo, N. The uses of somatic embryogenesis for genetic transformation. In *Somatic Embryogenesis: Fundamental Aspects and Applications*, 1st ed.; Loyola-Vargas, V., Ochoa-Alejo, N., Eds.; Springer International Publishing: Cham, Switzerland, 2016; pp. 415–434.
3. Ikeda, M.; Kamada, H. Comparison of molecular mechanisms of somatic and zygotic embryogenesis. In *Somatic Embryogenesis*, 1st ed.; Mujib, A., Samaj, J., Eds.; Springer: Berlin, Heidelberg, 2006; pp. 51–68.
4. Wójcikowska, B.; Gaj, M.D. Somatic embryogenesis in Arabidopsis. In *Somatic Embryogenesis: Fundamental Aspects and Applications*, 1st ed.; Loyola-Vargas, V., Ochoa-Alejo, N., Eds.; Springer International Publishing: Cham, Switzerland, 2016; pp. 185–199.
5. Gaj, M.D. Factors influencing somatic embryogenesis induction and plant regeneration with particular reference to *Arabidopsis thaliana* (L.) Heynh. *Plant Growth Regul.* **2004**, *43*, 27–47. [[CrossRef](#)]
6. Reinert, J. Morphogenese und ihre Kontrolle an Gewebekulturen aus Carotten. *Naturwissenschaften* **1958**, *45*, 344–345. [[CrossRef](#)]
7. Mujib, A. *Somatic Embryogenesis in Ornamentals and Its Applications*; Springer: New Delhi, India, 2016; pp. 67–86.
8. Serbent, M.P.; Rebelo, A.M.; Pinheiro, A.; Giongo, A.; Tavares, L.B.B. Biological agents for 2,4-dichlorophenoxyacetic acid herbicide degradation. *Appl. Microbiol. Biotechnol.* **2019**, *103*, 5065–5078. [[CrossRef](#)] [[PubMed](#)]
9. Teixeira, M.C.; Duque, P.; Sá-Correia, I. Environmental genomics: Mechanistic insights into toxicity of and resistance to the herbicide 2,4-D. *Trends Biotechnol.* **2007**, *25*, 363–370. [[CrossRef](#)]
10. Song, Y. Insight into the mode of action of 2,4-dichlorophenoxyacetic acid (2,4-D) as an herbicide. *J. Integr. Plant Biol.* **2014**, *56*, 106–113. [[CrossRef](#)]
11. Juárez-González, V.T.; López-Ruiz, B.A.; Baldrich, P.; Luján-Soto, E.; Meyers, B.C.; Dinkova, T.D. The explant developmental stage profoundly impacts small RNA-mediated regulation at the dedifferentiation step of maize somatic embryogenesis. *Sci. Rep.* **2019**, *9*. [[CrossRef](#)]
12. Delporte, F.; Muhovski, Y.; Pretova, A.; Watillon, B. Analysis of expression profiles of selected genes associated with the regenerative property and the receptivity to gene transfer during somatic embryogenesis in *Triticum aestivum* L. *Mol. Biol. Rep.* **2013**, *40*, 5883–5906. [[CrossRef](#)]
13. She, M.; Yin, G.; Li, J.; Li, X.; Du, L.; Ma, W.; Ye, X. Efficient regeneration potential is closely related to auxin exposure time and catalase metabolism during the somatic embryogenesis of immature embryos in *Triticum aestivum* L. *Mol. Biotechnol.* **2013**, *54*, 451–460. [[CrossRef](#)]
14. Ahmadpour, R.; Zare, N.; Asghari-zakarta, R.; Sheikhzadeh, P. Efficient in vitro somatic embryogenesis and plant regeneration from mature and immature embryos of wheat (*Triticum aestivum* L.). *Braz. Arch. Biol. Technol.* **2016**, *59*, 1–12. [[CrossRef](#)]
15. Gaj, M.D. Direct somatic embryogenesis as a rapid and efficient system for in vitro regeneration of *Arabidopsis thaliana*. *Plant Cell. Tissue Organ Cult.* **2001**, *64*, 39–46. [[CrossRef](#)]
16. Grossmann, K. Auxin herbicides: Current status of mechanism and mode of action. *Pest. Manag. Sci.* **2010**, *66*, 113–120. [[CrossRef](#)] [[PubMed](#)]
17. Eyer, L.; Vain, T.; Pařízková, B.; Oklestkova, J.; Barbez, E.; Kozubíková, H. 2,4-D and IAA amino acid conjugates show distinct metabolism in Arabidopsis. *PLoS ONE* **2016**, *11*, e0159269. [[CrossRef](#)] [[PubMed](#)]
18. De Rybel, B.; Audenaert, D.; Beeckman, T.; Kepinski, S. The past, present, and future of chemical biology in auxin research. *Acs Chem. Biol.* **2009**, *4*, 987–998. [[CrossRef](#)] [[PubMed](#)]
19. Teale, W.D.; Paponov, I.A.; Palme, K. Auxin in action: Signalling, transport and the control of plant growth and development. *Nat. Rev. Mol. Cell Biol.* **2006**, *7*, 847–859. [[CrossRef](#)]
20. Pufky, J.; Qiu, Y.; Rao, M.V.; Hurban, P.; Jones, A.M. The auxin-induced transcriptome for etiolated Arabidopsis seedlings using a structure/function approach. *Funct. Integr. Genom.* **2003**, *3*, 135–143.
21. Tan, X.; Calderon-Villalobos, L.I.A.; Sharon, M.; Zheng, C.; Robinson, C.V.; Estelle, M.; Zheng, N. Mechanism of auxin perception by the TIR1 ubiquitin ligase. *Nature* **2007**, *446*, 640–645. [[CrossRef](#)]
22. Woodward, A.W.; Bartel, B. Auxin: Regulation, action, and interaction. *Ann. Bot.* **2005**, *95*, 707–735. [[CrossRef](#)]
23. Hansen, H.; Grossmann, K. Auxin-induced ethylene triggers abscisic acid biosynthesis and growth inhibition. *Plant Physiol.* **2000**, *124*, 1437–1448. [[CrossRef](#)]
24. Arteca, R.N.; Arteca, J.M. Effects of brassinosteroid, auxin, and cytokinin on ethylene production in *Arabidopsis thaliana* plants. *J. Exp. Bot.* **2008**, *59*, 3019–3026. [[CrossRef](#)]



25. Pazmino, D.M.; Rodriguez-Serrano, M.; Romero-Puertas, M.C.; Archilla-Ruiz, A.; Del Rio, L.A.; Sandalio, L.M. Differential response of young and adult leaves to herbicide 2,4-dichlorophenoxyacetic acid in pea plants: Role of reactive oxygen species. *Plant Cell Environ.* **2011**, *34*, 1874–1889. [\[CrossRef\]](#)
26. Feher, A.; Pasternak, T.P.; Dudits, D. Transition of somatic plant cells to an embryogenic state. *Plant Cell. Tissue Organ Cult.* **2003**, *74*, 201–228. [\[CrossRef\]](#)
27. Zavattieri, M.A.; Frederico, A.M.; Lima, M.; Sabino, R.; Arnholdt-Schmitt, B. Induction of somatic embryogenesis as an example of stress-related plant reactions. *Electron. J. Biotechnol.* **2010**, *13*, 12–13. [\[CrossRef\]](#)
28. Thibaud-Nissen, F.; Shealy, R.T.; Khanna, A.; Vodkin, L.O. Clustering of microarray data reveals transcript patterns associated with somatic embryogenesis in soybean. *Plant Physiol.* **2003**, *132*, 118–136. [\[CrossRef\]](#) [\[PubMed\]](#)
29. Mantiri, F.R.; Kurdyukov, S.; Lohar, D.P.; Sharopova, N.; Saeed, N.A.; Wang, X.-D.; VandenBosch, K.A.; Rose, R.J. The Transcription Factor MtSERF1 of the ERF Subfamily Identified by Transcriptional Profiling Is Required for Somatic Embryogenesis Induced by Auxin Plus Cytokinin in *Medicago truncatula* [W][OA]. *Plant Physiol.* **2008**, *146*, 1622–1636. [\[CrossRef\]](#) [\[PubMed\]](#)
30. Sharma, S.K.; Millam, S.; Hedley, P.E.; McNicol, J.; Bryan, G.J. Molecular regulation of somatic embryogenesis in potato: An auxin led perspective. *Plant Mol. Biol.* **2008**, *68*, 185–201. [\[CrossRef\]](#)
31. Imin, N.; Goffard, N.; Nizamidin, M.; Rolfe, B.G. Genome-wide transcriptional analysis of super-embryogenic *Medicago truncatula* explant cultures. *BMC Plant Biol.* **2008**, *8*, 110. [\[CrossRef\]](#)
32. Lin, H.C.; Morcillo, F.; Dussert, S.; Tranchant-Dubreuil, C.; Tregear, J.W.; Tranbarger, T.J. Transcriptome analysis during somatic embryogenesis of the tropical monocot *Elaeis guineensis*: Evidence for conserved gene functions in early development. *Plant Mol. Biol.* **2009**, *70*, 173–192. [\[CrossRef\]](#)
33. Sun, L.; Wu, Y.; Su, S.; Liu, H.; Yang, G.; Li, S. Differential gene expression during somatic embryogenesis in the maize (*Zea mays* L.) inbred line H99. *Plant Cell. Tissue Organ Cult.* **2012**, *109*, 271–286. [\[CrossRef\]](#)
34. Shani, E.; Salehin, M.; Zhang, Y.; Sanchez, S.E.; Doherty, C.; Wang, R.; Mangado, C.C.; Song, L.; Tal, I.; Pisanty, O.; et al. Plant Stress Tolerance Requires Auxin-Sensitive Aux/IAA Transcriptional Repressors. *Curr. Biol.* **2017**, *27*, 437–444. [\[CrossRef\]](#)
35. Blakeslee, J.J.; Spatola Rossi, T.; Kriechbaumer, V. Auxin biosynthesis: Spatial regulation and adaptation to stress. *J. Exp. Bot.* **2019**, *70*, 5041–5049. [\[CrossRef\]](#)
36. Wójcikowska, B.; Jaskóła, K.; Gasiorek, P.; Meus, M.; Nowak, K.; Gaj, M.D. LEAFY COTYLEDON2 (LEC2) promotes embryogenic induction in somatic tissues of *Arabidopsis*, via YUCCA-mediated auxin biosynthesis. *Planta* **2013**, *238*, 425–440. [\[CrossRef\]](#)
37. Wójcikowska, B.; Gaj, M.D. Expression profiling of *AUXIN RESPONSE FACTOR* genes during somatic embryogenesis induction in *Arabidopsis*. *Plant Cell Rep.* **2017**, *36*, 843–858. [\[CrossRef\]](#)
38. Ivanova, A.; Velcheva, M.; Denchev, P.; Atanassov, A.; Van Onckelen, H.A. Endogenous hormone levels during direct somatic embryogenesis in *Medicago falcata*. *Physiol. Plant.* **1994**, *92*, 85–89. [\[CrossRef\]](#)
39. Guiderdoni, E.; Mérot, B.; Eksomtramage, T.; Paulet, F.; Feldmann, P.; Glaszmann, J.C. Somatic embryogenesis in sugarcane (*Saccharum* Species). In *Somatic Embryogenesis and Synthetic Seed II*, 1st ed.; Bajaj, Y.P.S., Ed.; Springer: Berlin, Heidelberg, 1995; Volume 31, pp. 92–113.
40. Michalczuk, L.; Druart, P. Indole-3-acetic acid metabolism in hormone-autotrophic, embryogenic callus of Inmil® cherry rootstock (*Prunus incisa* x *serrula* 'GM 9') and in hormone-dependent, nonembryogenic calli of *Prunus incisa* x *serrula* and *Prunus domestica*. *Physiol. Plant.* **1999**, *107*, 426–432. [\[CrossRef\]](#)
41. Jiménez, V.M.; Bangerth, F. Endogenous hormone levels in explants and in embryogenic and non-embryogenic cultures of carrot. *Physiol. Plant.* **2001**, *111*, 389–395. [\[CrossRef\]](#) [\[PubMed\]](#)
42. Jiménez, V.M.; Bangerth, F. Endogenous hormone concentrations and embryogenic callus development in wheat. *Plant Cell. Tissue Organ Cult.* **2001**, *67*, 37–46. [\[CrossRef\]](#)
43. Jiménez, V.M.; Bangerth, F. Hormonal status of maize initial explants and of the embryogenic and non-embryogenic callus cultures derived from them as related to morphogenesis in vitro. *Plant Sci.* **2001**, *160*, 247–257. [\[CrossRef\]](#)
44. Grzyb, M.; Kalandyk, A.; Waligórski, P.; Mikuła, A. The content of endogenous hormones and sugars in the process of early somatic embryogenesis in the tree fern *Cyathea delgadii* Sternb. *Plant Cell. Tissue Organ Cult.* **2017**, *129*, 387–397. [\[CrossRef\]](#)

45. Li, Q.; Deng, C.; Xia, Y.; Kong, L.; Zhang, H.; Zhang, S.; Wang, J. Identification of novel miRNAs and miRNA expression profiling in embryogenic tissues of *Picea balfouriana* treated by 6-benzylaminopurine. *PLoS ONE* **2017**, *12*, 1–17. [\[CrossRef\]](#)
46. Liao, Y.K.; Liao, C.K.; Ho, Y.L. Maturation of somatic embryos in two embryogenic cultures of *Picea morrissonicola* Hayata as affected by alternation of endogenous IAA content. *Plant Cell. Tissue Organ Cult.* **2008**, *93*, 257–268. [\[CrossRef\]](#)
47. Zhou, X.; Zheng, R.; Liu, G.; Xu, Y.; Zhou, Y.; Laux, T.; Zhen, Y.; Harding, S.A.; Shi, J.; Chen, J. Desiccation treatment and endogenous IAA levels are key factors influencing high frequency somatic embryogenesis in *Cunninghamia lanceolata* (Lamb.) Hook. *Front. Plant Sci.* **2017**, *8*, 2054. [\[CrossRef\]](#) [\[PubMed\]](#)
48. Michalczyk, L.; Cooke, T.J.; Cohen, J.D. Auxin levels at different stages of carrot somatic embryogenesis. *Phytochemistry* **1992**, *31*, 1097–1103. [\[CrossRef\]](#)
49. Thomas, C.; Bronner, R.; Molinier, J.; Prinsen, E.; Van Onckelen, H.; Hahne, G. Immuno-cytochemical localization of indole-3-acetic acid during induction of somatic embryogenesis in cultured sunflower embryos. *Planta* **2002**, *215*, 577–583. [\[CrossRef\]](#) [\[PubMed\]](#)
50. Ayil-Gutiérrez, B.; Galaz-Avalos, R.M.; Peña-Cabrera, E.; Loyola-Vargas, V.M. Dynamics of the concentration of IAA and some of its conjugates during the induction of somatic embryogenesis in *Coffea canephora*. *Plant Signal. Behav.* **2013**, *8*, e26998. [\[CrossRef\]](#)
51. Márquez-López, R.E.; Pérez-Hernández, C.; Ku-González, Á.; Galaz-Ávalos, R.M.; Loyola-Vargas, V.M. Localization and transport of indole-3-acetic acid during somatic embryogenesis in *Coffea canephora*. *Protoplasma* **2018**, *255*, 695–708. [\[CrossRef\]](#)
52. Awada, R.; Campa, C.; Gibault, E.; Déchamp, E.; Georget, F.; Lepelley, M.; Abdallah, C.; Erban, A.; Seidel, M.; Kopka, J.; et al. Unravelling the Metabolic and Hormonal Machinery During Key Steps of Somatic Embryogenesis: A Case Study in Coffee. *Int. J. Mol. Sci.* **2019**, *20*, 4665. [\[CrossRef\]](#)
53. Pasternak, T.P.; Prinsen, E.; Ayaydin, F.; Miskolczi, P.; Potters, G.; Asard, H.; Van Onckelen, H.A.; Dudits, D.; Fehér, A. The role of auxin, pH, and stress in the activation of embryogenic cell division in leaf protoplast-derived cells of alfalfa. *Plant Physiol.* **2002**, *129*, 1807–1819. [\[CrossRef\]](#)
54. Pescador, R.; Kerbaui, G.B.; De Ferreira, W.M.; Purgatto, E.; Suzuki, R.M.; Guerra, M.P. A hormonal misunderstanding in *Acca sellowiana* embryogenesis: Levels of zygotic embryogenesis do not match those of somatic embryogenesis. *Plant Growth Regul.* **2012**, *68*, 67–76. [\[CrossRef\]](#)
55. Cheng, W.H.; Zhu, H.G.; Tian, W.G.; Zhu, S.H.; Xiong, X.P.; Sun, Y.Q.; Zhu, Q.H.; Sun, J. De novo transcriptome analysis reveals insights into dynamic homeostasis regulation of somatic embryogenesis in upland cotton (*G. hirsutum* L.). *Plant Mol. Biol.* **2016**, *92*, 279–292. [\[CrossRef\]](#)
56. Vondrakova, Z.; Dobrev, P.I.; Pesek, B.; Fischerova, L.; Vagner, M.; Motyka, V. Profiles of endogenous phytohormones over the course of Norway spruce somatic embryogenesis. *Front. Plant Sci.* **2018**, *9*, 1283. [\[CrossRef\]](#) [\[PubMed\]](#)
57. Wójcikowska, B.; Gaj, M.D. LEAFY COTYLEDON2-mediated control of the endogenous hormone content: Implications for the induction of somatic embryogenesis in Arabidopsis. *Plant Cell. Tissue Organ Cult.* **2015**, *121*, 255–258. [\[CrossRef\]](#)
58. Ribnicky, D.M.; Cohen, J.D.; Hu, W.S.; Cooke, T.J. An auxin surge following fertilization in carrots: A mechanism for regulating plant totipotency. *Planta* **2002**, *214*, 505–509. [\[PubMed\]](#)
59. Robert, H.; Park, C.; Gutiérrez, C.L.; Wójcikowska, B.; Pěnčík, A.; Novák, O.; Chen, J.; Grunewald, W.; Dresselhaus, T.; Friml, J.; et al. Maternal auxin supply contributes to early embryo patterning in Arabidopsis. *Nat. Plants* **2018**, *4*, 548–553. [\[CrossRef\]](#)
60. Robert, H.S. Molecular communication for coordinated seed and fruit development: What can we learn from auxin and sugars? *Int. J. Mol. Sci.* **2019**, *20*, 936. [\[CrossRef\]](#)
61. Winkelmann, T. Somatic versus zygotic embryogenesis: Learning from seeds. In *In Vitro Embryogenesis in Higher Plants*, 1st ed.; Germana, M.A., Lambardi, M., Eds.; Springer Science+Business Media LLC: New York, NY, USA, 2016; pp. 25–46.
62. Zhao, Y. Auxin biosynthesis and its role in plant development. *Annu. Rev. Plant Biol.* **2010**, *61*, 49–64. [\[CrossRef\]](#)
63. Wang, B.; Chu, J.; Yu, T.; Xu, Q.; Sun, X.; Yuan, J.; Xiong, G.; Wang, G.; Wang, Y.; Li, J. Tryptophan-independent auxin biosynthesis contributes to early embryogenesis in Arabidopsis. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, 4821–4826. [\[CrossRef\]](#)

64. Wickramasuriya, A.M.; Dunwell, J.M. Global scale transcriptome analysis of Arabidopsis embryogenesis in vitro. *Bmc Genom.* **2015**, *16*, 301. [[CrossRef](#)]
65. Bai, B.; Su, Y.H.; Yuan, J.; Zhang, X.S. Induction of somatic embryos in Arabidopsis requires local YUCCA expression mediated by the down-regulation of ethylene biosynthesis. *Mol. Plant* **2013**, *6*, 1247–1260. [[CrossRef](#)]
66. Robert, H.S.; Crhak Khaitova, L.; Mroue, S.; Benková, E. The importance of localized auxin production for morphogenesis of reproductive organs and embryos in Arabidopsis. *J. Exp. Bot.* **2015**, *66*, 5029–5042. [[CrossRef](#)]
67. Radoeva, T.; Lokerse, A.S.; Llavata-Peris, C.I.; Wendrich, J.R.; Xiang, D.; Liao, C.Y.; Vlaar, L.; Boekschoten, M.; Hooiveld, G.; Datla, R.; et al. A robust auxin response network controls embryo and suspensor development through a basic helix loop helix transcriptional module. *Plant Cell* **2019**, *31*, 52–67. [[CrossRef](#)] [[PubMed](#)]
68. Wójcikowska, B.; Botor, M.; Morończyk, J.; Wójcik, A.M.; Nodzyński, T.; Karcz, J.; Gaj, M.D. Trichostatin A triggers an embryogenic transition in Arabidopsis explants via an auxin-related pathway. *Front. Plant Sci.* **2018**, *9*, 1353. [[CrossRef](#)] [[PubMed](#)]
69. Weijers, D.; Wagner, D. Transcriptional Responses to the Auxin Hormone. *Annu. Rev. Plant Biol.* **2016**, *67*, 539–574. [[CrossRef](#)] [[PubMed](#)]
70. Yang, X.; Zhang, X.; Yuan, D.; Jin, F.; Zhang, Y.; Xu, J. Transcript profiling reveals complex auxin signalling pathway and transcription regulation involved in dedifferentiation and redifferentiation during somatic embryogenesis in cotton. *Bmc Plant Biol.* **2012**, *12*, 110. [[CrossRef](#)] [[PubMed](#)]
71. Xu, Z.; Zhang, C.; Zhang, X.; Liu, C.; Wu, Z.; Yang, Z.; Zhou, K.; Yang, X.; Li, F. Transcriptome profiling reveals auxin and cytokinin regulating somatic embryogenesis in different sister lines of cotton cultivar CCRI24. *J. Integr. Plant Biol.* **2013**, *55*, 631–642. [[CrossRef](#)] [[PubMed](#)]
72. Indoliya, Y.; Tiwari, P.; Chauhan, A.S.; Goel, R.; Shri, M.; Bag, S.K.; Chakrabarty, D. Decoding regulatory landscape of somatic embryogenesis reveals differential regulatory networks between japonica and indica rice subspecies. *Sci. Rep.* **2016**, *6*, 23050. [[CrossRef](#)] [[PubMed](#)]
73. Saptari, R.T.; Susila, H. Data mining study of hormone biosynthesis gene expression reveals new aspects of somatic embryogenesis regulation. *Vitr. Cell. Dev. Biol.-Plant* **2019**, *55*, 139–152. [[CrossRef](#)]
74. Singla, B.; Tyagi, A.K.; Khurana, J.P.; Khurana, P. Analysis of expression profile of selected genes expressed during auxin-induced somatic embryogenesis in leaf base system of wheat (*Triticum aestivum*) and their possible interactions. *Plant Mol. Biol.* **2007**, *65*, 677–692. [[CrossRef](#)]
75. Cao, A.; Zheng, Y.; Yu, Y.; Wang, X.; Shao, D.; Sun, J.; Cui, B. Comparative transcriptome analysis of SE initial dedifferentiation in cotton of different SE capability. *Sci. Rep.* **2017**, *7*, 1–13. [[CrossRef](#)]
76. Gliwicka, M.; Nowak, K.; Balazadeh, S.; Mueller-Roeber, B.; Gaj, M.D. Extensive Modulation of the transcription factor transcriptome during somatic embryogenesis in *Arabidopsis thaliana*. *PLoS ONE* **2013**, *8*. [[CrossRef](#)]
77. Shi, X.; Zhang, C.; Liu, Q.; Zhang, Z.; Zheng, B.; Bao, M. De novo comparative transcriptome analysis provides new insights into sucrose induced somatic embryogenesis in camphor tree (*Cinnamomum camphora* L.). *Bmc Genom.* **2016**, *17*, 26. [[CrossRef](#)] [[PubMed](#)]
78. Fadryin, N.; Rohani, E.R.; Muhammed-Hussein, Z.A.; Noor, N.M. Somatic embryogenesis-related gene expression and functional genomics in mangosteen. *Plant Gene* **2018**, *15*, 51–66. [[CrossRef](#)]
79. Capote, T.; Usié, A.; Barbosa, P.; Ramos, M.; Morais-Cecílio, L.; Gonçalves, S. Transcriptome dynamics of cork oak (*Quercus suber*) somatic embryogenesis reveals active gene players in transcription regulation and phytohormone homeostasis of embryo development. *Tree Genet. Genomes* **2019**, *15*, 52. [[CrossRef](#)]
80. Liu, W.; Wang, C.; Shen, X.; Liang, H.; Wang, Y.; He, Z.; Zhang, D.; Chen, F. Comparative transcriptome analysis highlights the hormone effects on somatic embryogenesis in *Catalpa bungei*. *Plant Reprod.* **2019**, *32*, 141–151. [[CrossRef](#)] [[PubMed](#)]
81. Quintana-Escobar, A.O.; Nic-Can, G.I.; Avalos, R.M.G.; Loyola-Vargas, V.M.; Gongora-Castillo, E. Transcriptome analysis of the induction of somatic embryogenesis in *Coffea canephora* and the participation of ARF and Aux/IAA genes. *PeerJ* **2019**, *7*, e7752. [[CrossRef](#)] [[PubMed](#)]
82. Chen, M.; Zhang, J.; Zhou, Y.; Li, S.; Fan, X.; Yang, L.; Guan, Y.; Zhang, Y. Transcriptome analysis of Lilium Oriental × Trumpet hybrid roots reveals auxin-related genes and stress-related genes involved in picloram-induced somatic embryogenesis induction. *J. Hort. Sci. Biotechnol.* **2019**, *94*, 317–330. [[CrossRef](#)]
83. Leyser, O. Auxin signaling. *Plant Physiol.* **2018**, *176*, 465–479. [[CrossRef](#)]

84. Li, S.B.; Xie, Z.Z.; Hu, C.G.; Zhang, J.Z. A review of AUXIN RESPONSE FACTORS (ARFs) in plants. *Front. Plant Sci.* **2016**, *7*, 47. [\[CrossRef\]](#)
85. Luo, J.; Zhou, J.J.; Zhang, J.Z. Aux/IAA gene family in plants: Molecular structure, regulation, and function. *Int. J. Mol. Sci.* **2018**, *19*, 259. [\[CrossRef\]](#)
86. Zheng, Y.; Ren, N.; Wang, H.; Stromberg, A.J.; Perry, S.E. Global identification of targets of the Arabidopsis MADS domain protein AGAMOUS-like15. *Plant Cell* **2009**, *21*, 2563–2577. [\[CrossRef\]](#)
87. Rensing, S.A.; Lang, D.; Schumann, E.; Reski, R.; Hohe, A. EST sequencing from embryogenic *Cyclamen persicum* cell cultures identifies a high proportion of transcripts homologous to plant genes involved in somatic embryogenesis. *J. Plant Growth Regul.* **2005**, *24*, 102–115. [\[CrossRef\]](#)
88. Ooi, S.E.; Choo, C.N.; Ishak, Z.; Ong-Abdullah, M. A candidate auxin-responsive expression marker gene, *EgIAA9*, for somatic embryogenesis in oil palm (*Elaeis guineensis* Jacq.). *Plant Cell. Tissue Organ Cult.* **2012**, *110*, 201–212. [\[CrossRef\]](#)
89. Wójcik, A.M.; Gaj, M.D. miR393 contributes to the embryogenic transition induced in vitro in Arabidopsis via the modification of the tissue sensitivity to auxin treatment. *Planta* **2016**, *244*, 231–243. [\[CrossRef\]](#) [\[PubMed\]](#)
90. Yang, X.; Wang, L.; Yuan, D.; Lindsey, K.; Zhang, X. Small RNA and degradome sequencing reveal complex miRNA regulation during cotton somatic embryogenesis. *J. Exp. Bot.* **2013**, *64*, 1521–1536. [\[CrossRef\]](#) [\[PubMed\]](#)
91. Villalobos, L.I.A.C.; Lee, S.; De Oliveira, C.; Ivetac, A.; Brandt, W.; Armitage, L.; Sheard, L.B.; Tan, X.; Parry, G.; Mao, H.; et al. A combinatorial TIR1/AFB–Aux/IAA co-receptor system for differential sensing of auxin. *Nat. Chem. Biol.* **2012**, *8*, 477. [\[CrossRef\]](#) [\[PubMed\]](#)
92. Braybrook, S.A.; Stone, S.L.; Park, S.; Bui, A.Q.; Le, B.H.; Fischer, R.L.; Goldberg, R.B.; Harada, J.J. Genes directly regulated by LEAFY COTYLEDON2 provide insight into the control of embryo maturation and somatic embryogenesis. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 3468–3473. [\[CrossRef\]](#)
93. Tvorogova, V.E.; Lutova, L.A. Genetic regulation of zygotic embryogenesis in angiosperm plants. *Russ. J. Plant Physiol.* **2018**, *65*, 1–14. [\[CrossRef\]](#)
94. Tang, X.; Bian, S.; Tang, M.; Lu, Q.; Li, S.; Liu, X.; Tian, G.; Nguyen, V.; Tsang, E.W.T.; Wang, A.; et al. MicroRNA-mediated repression of the seed maturation program during vegetative development in Arabidopsis. *Plos Genet.* **2012**, *8*, 20–22. [\[CrossRef\]](#)
95. Wójcik, A.M.; Nodine, M.D.; Gaj, M.D. miR160 and miR166/165 contribute to the LEC2-mediated auxin response involved in the somatic embryogenesis induction in Arabidopsis. *Front. Plant Sci.* **2017**, *8*, 2024. [\[CrossRef\]](#)
96. Müller, C.J.; Valdés, A.E.; Wang, G.; Ramachandran, P.; Beste, L.; Uddenberg, D.; Carlsbecker, A. PHABULOSA mediates an auxin signaling loop to regulate vascular patterning in Arabidopsis. *Plant Physiol.* **2016**, *170*, 956–970. [\[CrossRef\]](#)
97. Möller, B.K.; Ten Hove, C.A.; Xiang, D.; Williams, N.; López, L.G.; Yoshida, S.; Smit, M.; Datla, R.; Weijers, D. Auxin response cell-autonomously controls ground tissue initiation in the early Arabidopsis embryo. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, E2533–E2539. [\[CrossRef\]](#)
98. Donner, T.J.; Sherr, I.; Scarpella, E. Regulation of preprocambial cell state acquisition by auxin signaling in Arabidopsis leaves. *Development* **2009**, *136*, 3235–3246. [\[CrossRef\]](#) [\[PubMed\]](#)
99. Schlereth, A.; Möller, B.; Liu, W.; Kientz, M.; Flipse, J.; Rademacher, E.H.; Schmid, M.; Jürgens, G.; Weijers, D. MONOPTEROS controls embryonic root initiation by regulating a mobile transcription factor. *Nature* **2010**, *464*, 913–916. [\[CrossRef\]](#)
100. Krogan, N.T.; Yin, X.; Ckurshumova, W.; Berleth, T. Distinct subclades of Aux/IAA genes are direct targets of ARF5/MP transcriptional regulation. *New Phytol.* **2014**, *204*, 474–483. [\[CrossRef\]](#)
101. Wenzel, C.L.; Schuetz, M.; Yu, Q.; Mattsson, J. Dynamics of MONOPTEROS and PIN-FORMED1 expression during leaf vein pattern formation in *Arabidopsis thaliana*. *Plant J.* **2007**, *49*, 387–398. [\[CrossRef\]](#)
102. Krogan, N.T.; Marcos, D.; Weiner, A.I.; Berleth, T. The auxin response factor MONOPTEROS controls meristem function and organogenesis in both the shoot and root through the direct regulation of PIN genes. *New Phytol.* **2016**, *212*, 42–50. [\[CrossRef\]](#)
103. Su, Y.H.; Zhao, X.Y.; Liu, Y.B.; Zhang, C.L.; O'Neill, S.D.; Zhang, X.S. Auxin-induced WUS expression is essential for embryonic stem cell renewal during somatic embryogenesis in Arabidopsis. *Plant J.* **2009**, *59*, 448–460. [\[CrossRef\]](#)



104. Hardtke, C.S.; Ckurshumova, W.; Vidaurre, D.P.; Singh, S.A.; Stamatiou, G.; Tiwari, S.B.; Hagen, G.; Gilfoyle, T.J.; Berleth, T. Overlapping and non-redundant functions of the Arabidopsis auxin response factors MONOPTEROS and NONPHOTOTROPIC HYPOCOTYL 4. *Development* **2004**, *131*, 1089–1100. [\[CrossRef\]](#)
105. Breuninger, H.; Rikirsch, E.; Hermann, M.; Ueda, M.; Laux, T. Differential expression of WOX genes mediates apical-basal axis formation in the Arabidopsis embryo. *Dev. Cell* **2008**, *14*, 867–876. [\[CrossRef\]](#)
106. Möller, B.; Weijers, D. Auxin control of embryo patterning. *Cold Spring Harb. Perspect. Biol.* **2009**, *1*, 1–13. [\[CrossRef\]](#)
107. Guan, C.; Wu, B.; Yu, T.; Wang, Q.; Krogan, N.T.; Liu, X.; Jiao, Y. Spatial auxin signaling controls leaf flattening in Arabidopsis. *Curr. Biol.* **2017**, *27*, 2940–2950. [\[CrossRef\]](#) [\[PubMed\]](#)
108. Gambino, G.; Minuto, M.; Boccacci, P.; Perrone, I.; Vallania, R.; Gribaudo, I. Characterization of expression dynamics of WOX homeodomain transcription factors during somatic embryogenesis in *Vitis vinifera*. *J. Exp. Bot.* **2010**, *62*, 1089–1101. [\[CrossRef\]](#) [\[PubMed\]](#)
109. Palovaara, J.; Hallberg, H.; Stasolla, C.; Hakman, I. Comparative expression pattern analysis of WUSCHEL-related homeobox 2 (WOX2) and WOX8/9 in developing seeds and somatic embryos of the gymnosperm *Picea abies*. *New Phytol.* **2010**, *188*, 122–135. [\[CrossRef\]](#) [\[PubMed\]](#)
110. Klimasewska, K.; Pelletier, G.; Overton, C.; Stewart, D.; Rutledge, R.G. Hormonally regulated overexpression of Arabidopsis WUS and conifer LEC1 (CHAP3A) in transgenic white spruce: Implications for somatic embryo development and somatic seedling growth. *Plant Cell Rep.* **2010**, *29*, 723–734. [\[CrossRef\]](#) [\[PubMed\]](#)
111. Rupps, A.; Raschke, J.; Rümmler, M.; Linke, B.; Zoglauer, K. Identification of putative homologs of *Larix decidua* to BABY BOOM (BBM), LEAFY COTYLEDON1 (LEC1), WUSCHEL-related HOMEBOX2 (WOX2) and SOMATIC EMBRYOGENESIS RECEPTOR-like KINASE (SERK) during somatic embryogenesis. *Planta* **2016**, *243*, 473–488. [\[CrossRef\]](#)
112. Tvorogova, V.E.; Lebedeva, M.A.; Lutova, L.A. Expression of WOX and PIN genes during somatic and zygotic embryogenesis in *Medicago truncatula*. *Russ. J. Genet.* **2015**, *51*, 1189–1198. [\[CrossRef\]](#)
113. Tvorogova, V.E.; Fedorova, Y.A.; Potsenkovskaya, E.A.; Kudriashov, A.A.; Efremova, E.P.; Kvitkovskaya, V.A.; Wolabu, T.W.; Zhang, F.; Tadege, M.; Lutova, L.A. The WUSCHEL-related homeobox transcription factor MtWOX9-1 stimulates somatic embryogenesis in *Medicago truncatula*. *Plant Cell. Tissue Organ Cult.* **2019**, *138*, 517–527. [\[CrossRef\]](#)
114. Chen, Y.; Xu, X.; Liu, Z.; Zhang, Z.; XuHan, X.; Lin, Y.; Lai, Z. Global scale transcriptome analysis reveals differentially expressed genes involve in early somatic embryogenesis in *Dimocarpus longan* Lour. *Bmc Genom.* **2020**, *21*, 1–22. [\[CrossRef\]](#)
115. Zhang, Z.; Tucker, E.; Hermann, M.; Laux, T. A molecular framework for the embryonic initiation of shoot meristem stem cells. *Dev. Cell* **2017**, *40*, 264–277. [\[CrossRef\]](#)
116. Qiao, M.; Zhao, Z.; Song, Y.; Liu, Z.; Cao, L.; Yu, Y.; Li, S.; Xiang, F. Proper regeneration from in vitro cultured *Arabidopsis thaliana* requires the microRNA-directed action of an auxin response factor. *Plant J.* **2012**, *71*, 14–22. [\[CrossRef\]](#)
117. Horstman, A.; Willemsen, V.; Boutilier, K.; Heidstra, R. AINTEGUMENTA-LIKE proteins: Hubs in a plethora of networks. *Trends Plant Sci.* **2014**, *19*, 146–157. [\[CrossRef\]](#) [\[PubMed\]](#)
118. Ding, Z.; Friml, J. Auxin regulates distal stem cell differentiation in Arabidopsis roots. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 12046–12051. [\[CrossRef\]](#) [\[PubMed\]](#)
119. Nowak, K.; Gaj, M.D. Transcription factors in regulations of somatic embryogenesis. In *Somatic Embryogenesis: Fundamental Aspects and Applications*, 1st ed.; Loyola-Vargas, V., Ochoa-Alejo, N., Eds.; Springer International Publishing: Cham, Switzerland, 2016; pp. 455–469.
120. Jia, H.; Suzuki, M.; McCarty, D.R. Regulation of the seed to seedling developmental phase transition by the LAFL and VAL transcription factor networks. *Wires Dev. Biol.* **2014**, *3*, 135–145. [\[CrossRef\]](#) [\[PubMed\]](#)
121. Horstman, A.; Li, M.; Heidmann, I.; Weemen, M.; Chen, B.; Muino, J.M.; Angenent, G.C.; Boutilier, K. The BABY BOOM transcription factor activates the LEC1-ABI3-FUS3-LEC2 network to induce somatic embryogenesis. *Plant Physiol.* **2017**, *175*, 848–857. [\[CrossRef\]](#)
122. Lotan, T.; Ohto, M.A.; Matsudaira Yee, K.; West, M.A.L.; Lo, R.; Kwong, R.W.; Yamagishi, K.; Fischer, R.L.; Goldberg, R.B.; Harada, J.J. Arabidopsis LEAFY COTYLEDON1 is sufficient to induce embryo development in vegetative cells. *Cell* **1998**, *93*, 1195–1205. [\[CrossRef\]](#)

123. Stone, S.L.; Kwong, L.W.; Yee, K.M.; Pelletier, J.; Lepiniec, L.; Fischer, R.L.; Goldberg, R.B.; Harada, J.J. LEAFY COTYLEDON2 encodes a B3 domain transcription factor that induces embryo development. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 11806–11811. [\[CrossRef\]](#)
124. Gaj, M.D.; Zhang, S.; Harada, J.J.; Lemaux, P.G. Leafy cotyledon genes are essential for induction of somatic embryogenesis of Arabidopsis. *Planta* **2005**, *222*, 977–988. [\[CrossRef\]](#)
125. Kurczyńska, E.U.; Gaj, M.D.; Ujczak, A.; Mazur, E. Histological analysis of direct somatic embryogenesis in *Arabidopsis thaliana* (L.) Heynh. *Planta* **2007**, *226*, 619–628. [\[CrossRef\]](#)
126. Ledwoń, A.; Gaj, M.D. LEAFY COTYLEDON2 gene expression and auxin treatment in relation to embryogenic capacity of Arabidopsis somatic cells. *Plant Cell Rep.* **2009**, *28*, 1677. [\[CrossRef\]](#)
127. Ledwoń, A.; Gaj, M.D. LEAFY COTYLEDON1, FUSCA3 expression and auxin treatment in relation to somatic embryogenesis induction in Arabidopsis. *Plant Growth Regul.* **2011**, *65*, 157–167. [\[CrossRef\]](#)
128. Belide, S.; Zhou, X.R.; Kennedy, Y.; Lester, G.; Shrestha, P.; Petrie, J.R.; Singh, S.P. Rapid expression and validation of seed-specific constructs in transgenic LEC2 induced somatic embryos of *Brassica napus*. *Plant Cell. Tissue Organ Cult.* **2013**, *113*, 543–553. [\[CrossRef\]](#)
129. Guo, F.; Liu, C.; Xia, H.; Bi, Y.; Zhao, C.; Zhao, S.; Hou, L.; Li, F.; Wang, X. Induced expression of AtLEC1 and AtLEC2 differentially promotes somatic embryogenesis in transgenic tobacco plants. *PLoS ONE* **2013**, *8*, e71714. [\[CrossRef\]](#) [\[PubMed\]](#)
130. Fister, A.S.; Landherr, L.; Perryman, M.; Zhang, Y.; Gultinan, M.J.; Maximova, S.N. Glucocorticoid receptor-regulated TcLEC2 expression triggers somatic embryogenesis in *Theobroma cacao* leaf tissue. *PLoS ONE* **2018**, *13*, 1–19. [\[CrossRef\]](#)
131. Brand, A.; Quimbaya, M.; Tohme, J.; Chavarriaga-Aguirre, P. Arabidopsis LEC1 and LEC2 orthologous genes are key regulators of somatic embryogenesis in cassava. *Front. Plant Sci.* **2019**, *10*, 673. [\[CrossRef\]](#)
132. Braybrook, S.; Harada, J. LECs go crazy in embryo development. *Trends Plant Sci.* **2008**, *13*, 624–630. [\[CrossRef\]](#)
133. Stone, S.L.; Braybrook, S.A.; Paula, S.L.; Kwong, L.W.; Meuser, J.; Pelletier, J.; Hsieh, T.F.; Fischer, R.L.; Goldberg, R.B.; Harada, J.J. Arabidopsis LEAFY COTYLEDON2 induces maturation traits and auxin activity: Implications for somatic embryogenesis. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 3151–3156. [\[CrossRef\]](#)
134. Junker, A.; Mönke, G.; Rutten, T.; Keilwagen, J.; Seifert, M.; Thi, T.M.N.; Renou, J.P.; Balzergue, S.; Viehöver, P.; Hähnel, U.; et al. Elongation-related functions of LEAFY COTYLEDON1 during the development of *Arabidopsis thaliana*. *Plant J.* **2012**, *71*, 427–442. [\[CrossRef\]](#)
135. Wang, H.; Caruso, L.V.; Downie, A.B.; Perry, S.E. The embryo MADS domain protein AGAMOUS-LIKE 15 directly regulates expression of a gene encoding an enzyme involved in gibberellin metabolism. *Plant Cell* **2004**, *16*, 1206–1219. [\[CrossRef\]](#)
136. Harding, E.W.; Tang, W.; Nichols, K.W.; Fernandez, D.E.; Perry, S.E. Expression and maintenance of embryogenic potential is enhanced through constitutive expression of AGAMOUS-Like 15. *Plant Physiol.* **2003**, *133*, 653–663. [\[CrossRef\]](#)
137. Zhu, C.; Perry, S.E. Control of expression and autoregulation of AGL15, a member of the MADS-box family. *Plant J.* **2005**, *41*, 583–594. [\[CrossRef\]](#)
138. Wang, F.; Perry, S.E. Identification of direct targets of FUSCA3, a key regulator of Arabidopsis seed development. *Plant Physiol.* **2013**, *161*, 1251–1264. [\[CrossRef\]](#) [\[PubMed\]](#)
139. Zheng, Q.; Perry, S.E. Alterations in the transcriptome of soybean in response to enhanced somatic embryogenesis promoted by orthologs of AGAMOUS-Like15 and AGAMOUS-Like18. *Plant Physiol.* **2014**, *164*, 1365–1377. [\[CrossRef\]](#) [\[PubMed\]](#)
140. Perry, S.E.; Zheng, Q.; Zheng, Y. Transcriptome analysis indicates that GmAGAMOUS-Like 15 may enhance somatic embryogenesis by promoting a dedifferentiated state. *Plant Signal. Behav.* **2016**, *11*, 1–7. [\[CrossRef\]](#) [\[PubMed\]](#)
141. Zheng, Q.; Zheng, Y.; Ji, H.; Burnie, W.; Perry, S.E. Gene regulation by the AGL15 transcription factor reveals hormone interactions in somatic embryogenesis. *Plant Physiol.* **2016**, *172*, 2374–2387. [\[CrossRef\]](#)
142. Aida, M.; Beis, D.; Heidstra, R.; Willemsen, V.; Blilou, I.; Galinha, C.; Nussaume, L.; Noh, Y.S.; Amasino, R.; Scheres, B. The PLETHORA genes mediate patterning of the Arabidopsis root stem cell niche. *Cell* **2004**, *119*, 109–120. [\[CrossRef\]](#)

143. Billou, I.; Xu, J.; Wildwater, M.; Willemssen, V.; Paponov, I.; Frimi, J.; Heldstra, R.; Aida, M.; Palme, K.; Scheres, B. The PIN auxin efflux facilitator network controls growth and patterning in Arabidopsis roots. *Nature* **2005**, *433*, 39–44. [[CrossRef](#)]
144. Boutilier, K.; Offringa, R.; Sharma, V.K.; Kieft, H.; Ouellet, T.; Zhang, L. Ectopic Expression of BABY BOOM triggers a conversion from vegetative to embryonic growth. *Plant Cell* **2002**, *14*, 1737–1749. [[CrossRef](#)]
145. Tsuwamoto, R.; Yokoi, S.; Takahata, Y. Arabidopsis EMBRYOMAKER encoding an AP2 domain transcription factor plays a key role in developmental change from vegetative to embryonic phase. *Plant Mol. Biol.* **2010**, *73*, 481–492. [[CrossRef](#)]
146. Szczygieł-Sommer, A.; Gaj, M.D. The miR396–GRF regulatory module controls the embryogenic response in Arabidopsis via an auxin-related pathway. *Int. J. Mol. Sci.* **2019**, *20*, 5221. [[CrossRef](#)]
147. Horstman, A.; Bemer, M.; Boutilier, K. A transcriptional view on somatic embryogenesis. *Regeneration* **2017**, *4*, 201–216. [[CrossRef](#)]
148. Passarinho, P.; Ketelaar, T.; Xing, M.; Van Arkel, J.; Maliepaard, C.; Hendriks, M.W.; Joosen, R.; Lammers, M.; Herdies, L.; Den Boer, B.; et al. BABY BOOM target genes provide diverse entry points into cell proliferation and cell growth pathways. *Plant Mol. Biol.* **2008**, *68*, 225–237. [[CrossRef](#)]
149. Pinon, V.; Prasad, K.; Grigg, S.P.; Sanchez-Perez, G.F.; Scheres, B. Local auxin biosynthesis regulation by PLETHORA transcription factors controls phyllotaxis in Arabidopsis. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 1107–1112. [[CrossRef](#)] [[PubMed](#)]
150. Ogas, J.; Kaufmann, S.; Henderson, J.; Somerville, C. PICKLE is a CHD3 chromatin-remodeling factor that regulates the transition from embryonic to vegetative development in Arabidopsis. *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 13839–13844. [[CrossRef](#)] [[PubMed](#)]
151. Zhang, H.; Rider, S.D.; Henderson, J.T.; Fountain, M.; Chuang, K.; Kandachar, V.; Simons, A.; Edenberg, H.J.; Romero-Severson, J.; Muir, W.M.; et al. The CHD3 remodeler PICKLE promotes trimethylation of histone H3 lysine 27. *J. Biol. Chem.* **2008**, *283*, 22637–22648. [[CrossRef](#)] [[PubMed](#)]
152. Suzuki, M.; Wang, H.H.Y.; McCarty, D.R. Repression of the Leafy Cotyledon 1/B3 regulatory network in plant embryo development by VP1/Abcisic Acid Insensitive 3-Like B3 genes. *Plant Physiol.* **2007**, *143*, 902–911. [[CrossRef](#)] [[PubMed](#)]
153. Zhou, Y.; Tan, B.; Luo, M.; Li, Y.; Liu, C.; Chen, C.; Yu, C.W.; Yang, S.; Dong, S.; Ruan, J.; et al. HISTONE DEACETYLASE19 interacts with HSL1 and participates in the repression of seed maturation genes in Arabidopsis seedlings. *Plant Cell* **2013**, *25*, 134–148. [[CrossRef](#)] [[PubMed](#)]
154. Chhun, T.; Chong, S.Y.; Park, B.S.; Wong, E.C.C.; Yin, J.L.; Kim, M.; Chua, N.H. HSI2 repressor recruits MED13 and HDA6 to down-regulate seed maturation gene expression directly during arabidopsis early seedling growth. *Plant Cell Physiol.* **2016**, *57*, 1689–1706. [[CrossRef](#)]
155. Van Der Graaff, E.; Laux, T.; Rensing, S.A. Protein family review The WUS homeobox-containing (WOX) protein family. *Genome Biol.* **2009**, *10*, 248. [[CrossRef](#)]
156. Dolzblasz, A.; Nardmann, J.; Clerici, E.; Causier, B.; Van der Graaff, E.; Chen, J.; Davies, B.; Werr, W.; Laux, T. Stem cell regulation by Arabidopsis WOX genes. *Mol. Plant* **2016**, *9*, 1028–1039. [[CrossRef](#)]
157. Zuo, J.; Niu, Q.W.; Frugis, G.; Chua, N.H. The WUSCHEL gene promotes vegetative-to-embryonic transition in Arabidopsis. *Plant J.* **2002**, *30*, 349–359. [[CrossRef](#)] [[PubMed](#)]
158. Solís-Ramos, L.Y.; González-Estrada, T.; Nahuath-Dzib, S.; Zapata-Rodriguez, L.C.; Castaño, E. Overexpression of WUSCHEL in *C. chinense* causes ectopic morphogenesis. *Plant Cell. Tissue Organ Cult.* **2009**, *96*, 279–287. [[CrossRef](#)]
159. Arroyo-Herrera, A.; Ku Gonzalez, A.; Canche Moo, R.; Quiroz-Figueroa, F.R.; Loyola-Vargas, V.M.; Rodriguez-Zapata, L.C.; Burgeff D'Hondt, C.; Suárez-Solís, V.M.; Castaño, E. Expression of WUSCHEL in *Coffea canephora* causes ectopic morphogenesis and increases somatic embryogenesis. *Plant Cell. Tissue Organ Cult.* **2008**, *94*, 171–180. [[CrossRef](#)]
160. Zheng, W.; Zhang, X.; Yang, Z.; Wu, J.; Li, F.; Duan, L.; Liu, C.; Lu, L.; Zhang, C.; Li, F. AtWuschel promotes formation of the embryogenic callus in *Gossypium hirsutum*. *PLoS ONE* **2014**, *9*, 1–8. [[CrossRef](#)]
161. Bouchabké-Coussa, O.; Obellianne, M.; Linderme, D.; Montes, E.; Maia-Grondard, A.; Vilaine, F.; Pannetier, C. WUSCHEL overexpression promotes somatic embryogenesis and induces organogenesis in cotton (*Gossypium hirsutum* L.) tissues cultured in vitro. *Plant Cell Rep.* **2013**, *32*, 675–686. [[CrossRef](#)] [[PubMed](#)]

162. Ma, Y.; Miotk, A.; Šutiković, Z.; Ermakova, O.; Wenzl, C.; Medzihradsky, A.; Gaillochet, C.; Forner, J.; Utan, G.; Brackmann, K.; et al. WUSCHEL acts as an auxin response rheostat to maintain apical stem cells in Arabidopsis. *Nat. Commun.* **2019**, *10*, 1–11. [\[CrossRef\]](#)
163. Magnani, E.; Jiménez-Gómez, J.M.; Soubigou-Taconnat, L.; Lepiniec, L.; Fiume, E. Profiling the onset of somatic embryogenesis in Arabidopsis. *Bmc Genom.* **2017**, *18*, 1–12. [\[CrossRef\]](#)
164. To, A.; Valon, C.; Savino, G.; Guillemot, J.; Devic, M.; Giraudat, J.; Parcy, F.A. network of local and redundant gene regulation governs Arabidopsis seed maturation. *Plant Cell* **2006**, *18*, 1642–1651. [\[CrossRef\]](#)
165. Kagaya, Y.; Toyoshima, R.; Okuda, R.; Usui, H.; Yamamoto, A.; Hattori, T. LEAFY COTYLEDON1 controls seed storage protein genes through its regulation of FUSCA3 and ABSCISIC ACID INSENSITIVE3. *Plant Cell Physiol.* **2005**, *46*, 399–406. [\[CrossRef\]](#)
166. Yamamoto, A.; Kagaya, Y.; Usui, H.; Hobo, T.; Takeda, S.; Hattori, T. Diverse roles and mechanisms of gene regulation by the Arabidopsis seed maturation master regulator FUS3 revealed by microarray analysis. *Plant Cell Physiol.* **2010**, *51*, 2031–2046. [\[CrossRef\]](#)
167. Gazzarrini, S.; Tsuchiya, Y.; Lumba, S.; Okamoto, M.; McCourt, P. The transcription factor FUSCA3 controls developmental timing in Arabidopsis through the hormones gibberellin and abscisic acid. *Dev. Cell* **2004**, *7*, 373–385. [\[CrossRef\]](#)
168. Hu, Y.; Zhou, L.; Huang, M.; He, X.; Yang, Y.; Liu, X.; Li, Y.; Hou, X. Gibberellins play an essential role in late embryogenesis of Arabidopsis. *Nat. Plants* **2018**, *4*, 289–298. [\[CrossRef\]](#) [\[PubMed\]](#)
169. Mu, J.; Tan, H.; Hong, S.; Liang, Y.; Zuo, J. Arabidopsis transcription factor genes NF-YA1, 5, 6, and 9 play redundant roles in male gametogenesis, embryogenesis, and seed development. *Mol. Plant* **2013**, *6*, 188–201. [\[CrossRef\]](#) [\[PubMed\]](#)
170. Hofmann, F.; Schon, M.A.; Nodine, M.D. The embryonic transcriptome of *Arabidopsis thaliana*. *Plant Reprod.* **2019**, *32*, 77–91. [\[CrossRef\]](#) [\[PubMed\]](#)
171. Rodrigues, A.S.; Chaves, I.; Costa, B.V.; Lin, Y.C.; Lopes, S.; Milhinhos, A.; Van de Peer, Y.; Miguel, C.M. Small RNA profiling in *Pinus pinaster* reveals the transcriptome of developing seeds and highlights differences between zygotic and somatic embryos. *Sci. Rep.* **2019**, *9*, 1–14. [\[CrossRef\]](#)



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).